# A Needle in a Haystack or Systematic Search for IncRNA Targets

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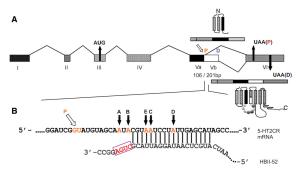
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#### Long non-coding RNAs

- ullet  $\simeq 40\%$  of the entire human genome is transcribed
- ullet  $\simeq 18\%$  of intergenic space is transcribed, generally at lower levels
- Some IncRNAs are involved in epigenetic silencing and imprinting
- Function most long non-coding transcripts yet unknown
- Diverse class of molecules with distinct functions

- Are there specific motifs in IncRNAs that are responsible for targeting to specific genomic loci?
- Do IncRNAs directly interact with DNA to form IncRNA:DNA hybrids or triplexes?
- If the specificity of lncRNAs is achieved by sequence complementarity, do they directly interact with other RNAs?
- Are there any IncRNAs implicated in the regulation of alternative splicing?

## snoRNA HBII-52 regulates splicing of 5- $\mathrm{HT_{2C}R}$ exon V



- Exon V has two donor sites, proximal (P) and distal (D)
- A truncated protein is produced when P is used
- HBII-52, a brain specific C/D box snoRNA, serves as a patch base-pairing to a sequence downstream of P
- $\bullet~$  HBII-52 and 5- $\rm HT_{\rm 2C}R$  are on different chromosomes
- HBII-52 also affects splicing of at least five other genes<sup>1</sup>

<sup>&</sup>lt;sup>1</sup>S. Kishore and S. Stamm, Science 311 no. 5758 pp. 230-232, 2006.

### Can we discover HBII-52 targets bioinformatically?

- BLAST or better GUUGle<sup>2</sup> (suffix trees + GU bps)
- ullet Blasting SNORD115 against all human genes gives  $\simeq 2,500$  hits
- ullet After filtering out snoRNA paralogs,  $\simeq 500$  hits left
- Other HBII-52 targets are imperfect, need internal loops

```
5' G C G 3' 5' A A G 3' 5' C A C 3'
GUGAUUCU UUG GUAU GUAU GCA CGUA UAAC CGU
CAUUAGGA AAC CGUA CGUA CGU
CA UAGGAUA ACUC UA GCAU GGA CGU
U U U 5' 3' G U C 5' 3' A U A 5'
```

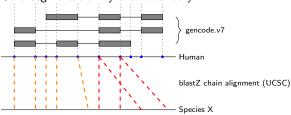
- Have internal loops? Sorry, no BLAST or GUUGle (but RNAplex3).
- There is an emerging need for a computational method that would allow efficient detection of RNA-RNA interaction sites on transcriptome-wide scale
- Conservation is a powerful and restrictive filter to narrow down the search to phylogenetically conserved interactions.

<sup>&</sup>lt;sup>2</sup>Gerlach & Giegerich, Bioinformatics, 22(6):762-764, 2006

<sup>&</sup>lt;sup>3</sup>Tafer *et al*, Bioinformatics 27(14):1934-40, 2011

#### Methods

Gene segmentation by exon boundary:



**IRBIS** 

- Sequence weights from phylogenetic tree (16 mammals)
- IRBIS
  - Set A = segments of non-coding genes (e.g., snoRNA, IncRNAs etc)
  - Set B = non-coding segments of protein-coding genes
  - $\triangleright$   $\mathcal{R} = A \times B$  (all-to-all)
  - ▶ Pattern 4-2-4, at most 1 GT and at least 2 GC per seed
  - Low-comlexity regions excluded
  - ▶ Present in 75% of species
  - ► Length at least 12 after extention

#### Gallery: intERmolecular structures

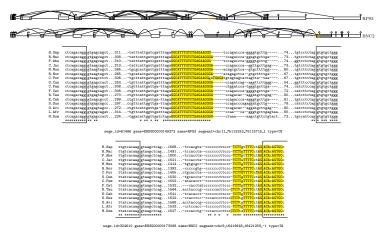
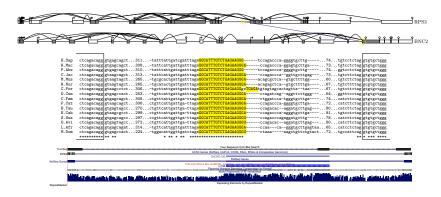


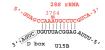


Figure 24: CS=21.87

#### Conserved box in RPS3 intron is a snoRNA



U15B is predicted to guide the 2'O-ribose methylation of 28S rRNA



- Why conservation extends beyond D-box?
- U15B is complementary to 11 other targets

#### HBII-52 and splicing of 5- $\mathrm{HT_{2C}R}$ exon V

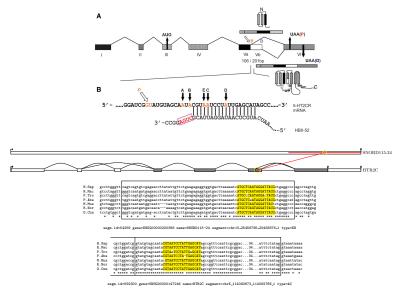


Figure 1: CS=

### Confounding factors

- There are reasons for a pair of motifs to be complementary and conserved other than RNA secondary structure
- Conserved bi-directional cis-elements on the DNA will always be found as such
- We can't distinguish them from conserved RNA-RNA interaction sites in principle
- (Sense-antisense pairs have to be excluded forever)

### Control 1: Search the opposite strand

- A = segments of IncRNAs
- B = (intronic) segments of protein coding genes
- Search A vs. B', sequences on the opposite strand to ones in B
- Conservation rate and dinucleotide content don't change

### Control 1: Search the opposite strand

In the tables: #hits[A, B]/#hits[A, B'] (% enrichment)

#### 16 placental mammals

	snoRNA	snRNA	IncRNA	introns
snoRNA		3/0 (NA)	277/241 (+14%)	1439/1099 (+30%)
snRNA			15/2 (NA)	120/92 (+30%)
IncRNA				7974/6329 (+25%)
introns				

#### 12 drosophilids

	snoRNA	snRNA	ncRNA	introns
snoRNA		71/95 (-25%)	34/39 (-12%)	1158/1122 (+3%)
snRNA			60/175 (-65%)	3432/2695 (+27%)
ncRNA				963/921 (+4%)
introns				

#### 6 nematodes

	snoRNA	snRNA	ncRNA	introns
snoRNA		107/69 (+55%)	514/512 (0%)	362/355 (+1%)
snRNA			2273/2584 (-12%)	1088/1117 (-2%)
ncRNA				5635/4950 (+13%)
introns				

Non-coding RNAs have higher potential to basepair introns of protein-coding genes at sense strand compared to antisense strand

### Control 2: Random sampling

- A = segments of IncRNAs
- Search set  $A_1$  vs. set B, where A is sampled randomly from "non-lncRNAs"
- $A_1$  = segments of protein coding genes (equivalent random sample)
- $\bullet$   $B_1 = B \setminus A_1$
- Search A against  $B_1$  against  $A_1$  against  $B_1$
- Conservation rate and GC content of the random sample are confounding
- How enrichment in A against  $B_1$  vs. A against  $B'_1$  relates to the enrichment  $A_1$ against  $B_1$  vs. A against  $B'_1$

## Control 2: Random sampling

- A =segments of lncRNAs
- $\bullet$  B = intronic segments of protein coding genes
- $A_1$  = random sample of segments of protein coding genes
- $B_1 = B \setminus A_1$
- $B_1'$  = reverse complements to sequences in  $B_1$

i	$\#[A, B_1]$	$\#[A, B_1']$	%	$\#[A_1, B_1]$	$\#[A_1, B_1']$	%	% - %
1	6653	5126	29.79	8482	7790	8.88	20.91
2	6756	5329	26.78	8166	7673	6.43	20.35
3	6661	5354	24.41	7922	7753	2.18	22.23
4	6581	5268	24.92	8864	8370	5.90	19.02
		***			***		
20	6737	5218	29.11	8252	7566	9.07	20.04

- Long non-coding RNAs have higher potential to basepair introns of protein-coding genes than do protein-coding genes themselves
- True in mammals (+20%), drosophilids (+7%), and nematodes (+15%)

#### Summary

- Non-coding RNAs have higher potential to basepair introns of protein-coding genes at sense strand as compared to antisense strand
- IncRNAs have higher potential to basepair introns of protein-coding genes than do protein-coding genes themselves
- IncRNAs predicted to be complementary to introns of protein-coding genes are, on average, more correlated (by absolute value) with the respective splicing events than do mock target pairs
- In spite of statistical evidence, we still don't know which pairs are functional

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**Postdocs wanted** 

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and in C. Notredame's lab