

# Patterns of tRNA Modifications

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Joint work with

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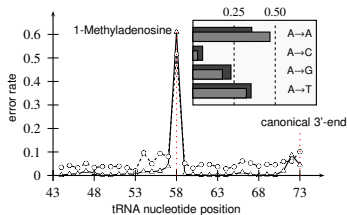
Benasque, Jul 17 2018

# Conservation of Modification Patterns

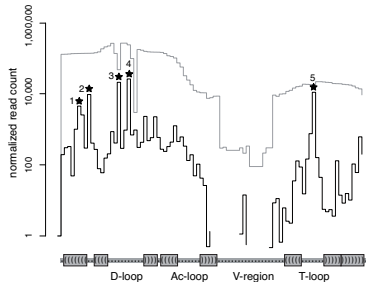
- How conserved are chemical modifications of RNAs
  - between different species
  - between different tissues
- Can this be studied with widely available data?  
tRNAs (or at least fragments) appear in high coverage in most small-RNA-seq datasets.  
Can this information be mined?

The answer is a partial yes: Many but not all chemical modification leave detectable traces in normal RNA-seq data

# Hallmarks of Modifications in RNA-seq data



mismatches



RT stops

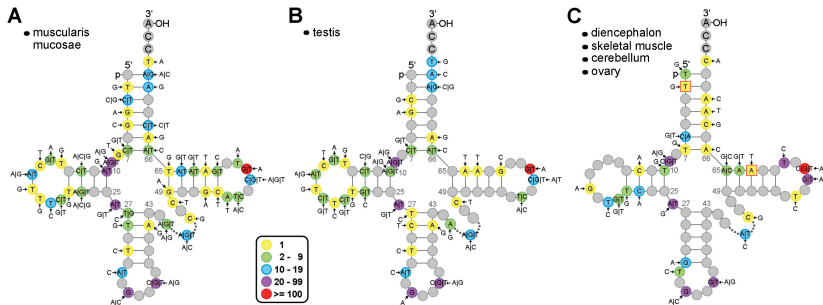
# Mapping of tRNA reads

- Many very similar tRNA paralog
- tRNAs with introns

## Specialized mapping pipeline

- remove pre-tRNA reads that map across the boundaries of mature tRNAs (and everything that maps well outside tRNAs)
- mask out tRNA genes from genome
- cluster very similar mature tRNAs and use a consensus
- add these “tRNA prototypes” as artificial chromosomes
- disregard positions with variations between paralogs in the same cluster

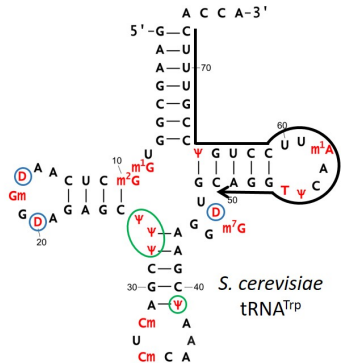
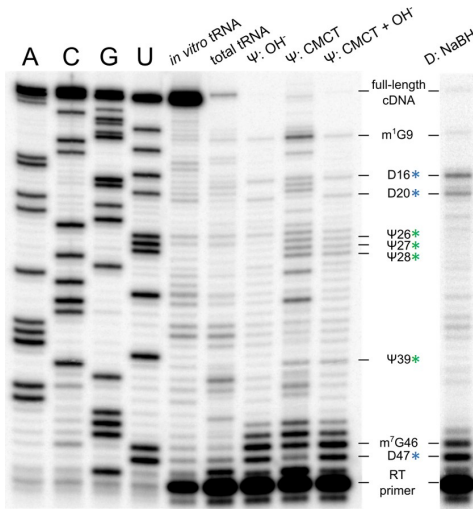
# Variations between tissues



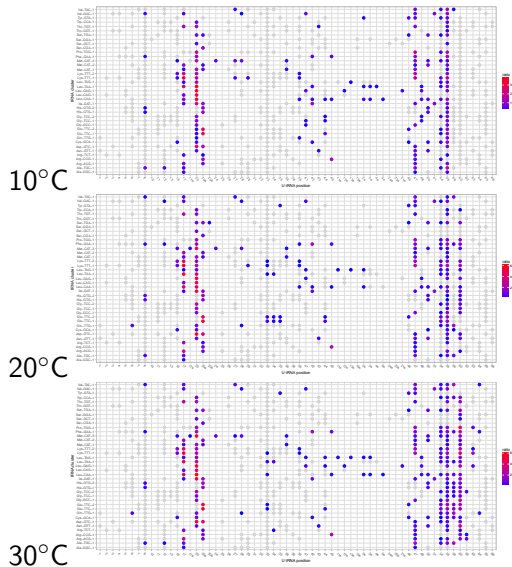
## More Results

- Very similar results for rat tissues
- Evidence for a few modifications in miRNAs
- the method also picks up A-I editing in miRNAs (and other small RNAs)
- modifications seem to be common also in snoRNAs

# Many modifications require chemical treatment



# Temperature dependence of Dihydrouridine

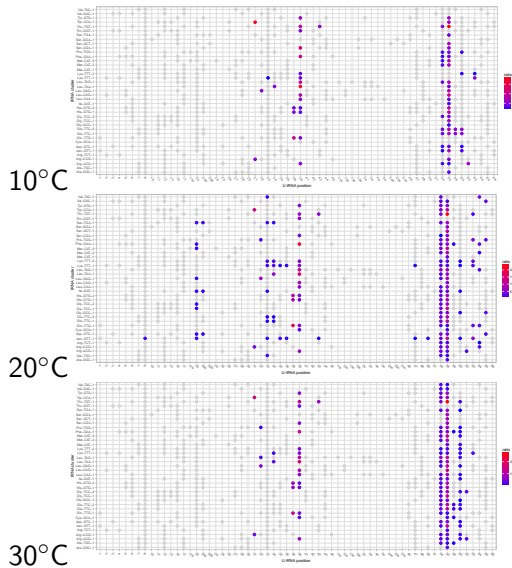


## *Exiguobacterium sibiricum*

- 3 biological replicates
- simple model to determine a  $p$ -value for the over-representation of RT-stops at each position
- Benjamini-Hochberg correction with 5% FDR
- cutoff for minimum fold change



# Temperature dependence of Pseudouridine



*Exiguobacterium sibiricum*