BGSU-NDB Collaboration: RNA Databases Update

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Benasque
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RNA.bgsu.edu ndbserver.rutgers.edu/





Acknowledgements

Craig Zirbel



Sri Appasamy



Jamie Cannone



BGSU



Dr. Anton Petrov Dr. Blake Sweeney



Funding:



Collaboration with Nucleic Acid Database



A Portal for Three-dimensional Structural Information about Nucleic Acids
As of 17-Apr-2018 number of released structures: 9452

Search DNA

Search RNA

Advanced Search

Enter an NDB ID or PDB ID

Search for released structures

Q

NDB Members

Goal

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Publications

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About the NDB

NDB Members

Co-founders:

Helen M. Berman, Ph.D. Rutgers University Wilma K. Olson, Ph.D. Rutgers University David Beveridge, Ph.D. Wesleyan University

Current Collaborators:

Neocles B. Leontis, Ph.D. Bowling Green state University Craig L. Zirbel, Ph.D. Bowling Green state University

Current NDB members:

Helen M. Berman, Ph.D. Rutgers University John Westbrook, Ph.D. Rutgers University Catherine L. Lawson, Ph.D. Rutgers University Brinda Vallat, Ph.D. Rutgers University Li Chen, Rutgers University Prof. Helen Berman



Dr. Cathy Lawson



The Goal

The goal of the Nucleic Acid Database Project (NDB) is to archive and distribute structural information about nucleic acids. The NDB was founded in 1992 by Helen M. Berman, Rutgers University, Wilma K. Olson, Rutgers University, and David Beveridge, Wesleyan University.

The NDB Project is funded by the National Institutes of Health and has been funded by National Science Foundation and the Department of Energy in the past.

Weekly annotation pipeline run on all RNA-containing 3D structures in PDB:

- Grouping structures based on Seq and 3D similarity
- 2. Annotation of RNA Interactions
- 3. Identification of 3D Motifs

Release id	All changes	Date	Number of IFEs
3.21 (current)	7 CHANGES	2018-05-11	8411
3.20	8 CHANGES	2018-05-04	8401
3.19	6 CHANGES	2018-04-27	8371
3.18	11 CHANGES	2018-04-20	8352
3.17	2 CHANGES	2018-04-13	8333
3.16	0 CHANGES	2018-04-06	8331
3.15	12 CHANGES	2018-03-29	8331
3.14	13 CHANGES	2018-03-22	8311
3.13	10 CHANGES	2018-03-15	8290
3.12	8 CHANGES	2018-03-08	8276
3.11	8 CHANGES	2018-03-01	8241
3.10	19 CHANGES	2018-02-23	8225
3.9	51 CHANGES	2018-02-16	8179
3.8	0 CHANGES	2018-02-09	8040

rna.bgsu.edu Representative sets

... running since 2011

Now based on mmCIF files:

IFE = Integrated Functional

Element: All RNA

chains that form 2D structure together

Ex: Eukaryal LSU

rRNA: 28S+5.8S

Release id	All changes	Date	Number of IFEs
0.15		2011-04-30	1699
0.14	4 CHANGES	2011-04-23	1699
0.13		2011-04-16	1696
0.12	3 CHANGES	2011-04-11	1696
0.11	12 CHANGES	2011-04-09	1696
0.10		2011-04-02	1678
0.9	3 CHANGES	2011-03-26	1677
0.8		2011-03-19	1670
0.7	3 CHANGES	2011-03-12	1670
0.6		2011-03-05	1652
0.5	2 CHANGES	2011-02-26	1650
0.4		2011-02-19	1642
0.3		2011-02-16	1642
0.2	3 CHANGES	2011-02-12	1639
0.1		2011-02-05	1630

rna.bgsu.edu Representative sets

Equivalence classes - same molecule, same organism

#	Equivalence class	Representative	Resolution	Nts	Class members
10	NR_4.0_75645.1 EXACT MATCH Toxoplasma gondii	5XXB 1 1+5XXB 1 4 (5XXB) • 25S RNA, 5.8S RNA • ELECTRON MICROSCOPY • Chain(s): 1, 4; model(s): 1	3.2 Å	3077	(1) 5XXB 1 1+5XXB 1 4
11	NR_4.0_41610.3 EXACT MATCH Bacillus subtilis	5NJT 1 U (5NJT) • 23S ribosomal RNA • ELECTRON MICROSCOPY • Chain(s): U; model(s): 1	3.8 Å	2923	(2) 5NJT 1 U, 3J9W 1 BA
12	NR_4.0_83717.35 EXACT MATCH Escherichia coli	5J7L 1 DA (5J7L) • 23S rRNA, 23S ribosomal RNA, RNA (2903-M[] • X-RAY DIFFRACTION • Chain(s): DA; model(s): 1	3.0 Å	2886	(146) 5J7L 1 DA, 5J8A 1 DA, 5JC9 1 DA, 4YBB 1 DA, 5 T8 1 DA, 4WO 1 BA, 4WO 1 CA, 5J91 1 DA, 4V9P 1 EA, 5J88 1 DA 4V9P 1 CA, 4V9P 1 GA, 4V9P 1 AA, 4V9D 1 CA, 4V9O 1 AA, 4V9O 1 EA, 4WWW 1 RA, 4V9O 1 CA, 4V6C 1 BA, 5J7L 1 CA, 4U27 1 BA, 4V7T 1 BA, 4WF1 1 BA, 5J8A 1 CA, 5JC9 1 CA,

Where? rna.bgsu.edu/rna3dhub/nrlist/release/3.21

Filter by organism; SSU's separate from LSU's

Filter: scrofa Showing 1 to 7 of 7 entries (filtered from 1,509 total entries)

#	Equivalence class	Representative	Resolution	Nts	Class members
3	NR_4.0_26483.1 EXACT MATCH Sus scrofa	3J7Q 1 5+3J7Q 1 8 (3J7Q) • 28S ribosomal RNA, 5.8S ribosomal RNA • ELECTRON MICROSCOPY • Chain(s): 5, 8; model(s): 1	3.4 Å	3662	(4) 3J7Q 1 5+3J7Q 1 8, 3J7O 1 5+3J7O 1 8, 3J7P 1 5+3J7P 1 8, 3J7R 1 5+3J7R 1 8
24	NR_4.0_61505.1 EXACT MATCH Sus scrofa	3J7P 1 S2 (3J7P) • 18S ribosomal RNA • ELECTRON MICROSCOPY • Chain(s): S2; model(s): 1	3.5 Å	1742	(2) 3J7P 1 S2, 3J7R 1 S2
35	NR_4.0_06715.2 EXACT MATCH Sus scrofa	4V19 1 A (4V19) • MITORIBOSOMAL 16S RRNA, TRNA, MITORIBOSO[] • ELECTRON MICROSCOPY • Chain(s): A; model(s): 1	3.4 Å	1515	(2) <u>4V19 1 A</u> , <u>5AJ4 1 BA</u>

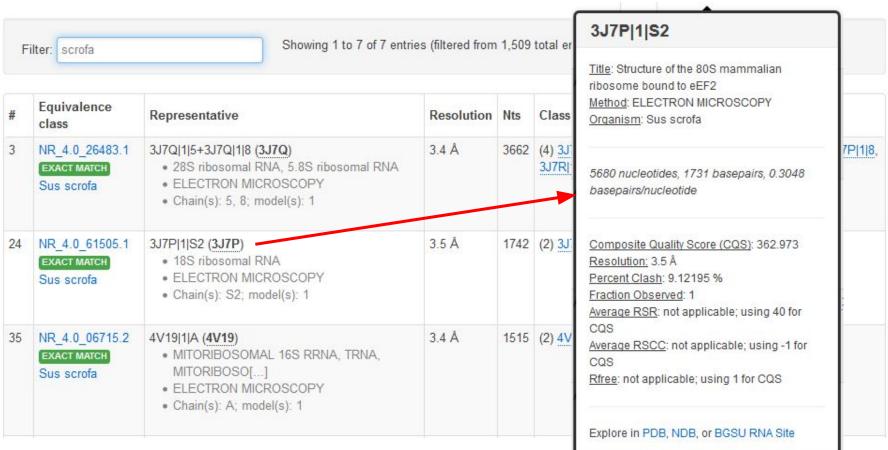
Composite Quality Score (CQS)

The representative for each equivalence class (EC) is chosen, starting with Release 3.0, as the Integrated Functional Element (IFE) with the optimal composite quality score (CQS). The CQS is the weighted sum of six components; five are defined at the IFE level, and one (Fraction Unobserved) is a property of the EC. The weights were developed to select the structure with the best experimental evidence for its reported coordinates as the EC representative.

CQS = (Resolution * 1.0) + (Percent Clash * 0.6) + (Average RSR * 8.0) + $((1 - \text{Average RSCC}) * 8.0) + (R_{\text{free}} * 18.0) + (Fraction Unobserved * 4.0)$

[Percent Clash is the percentage of nucleotides having steric clashes; Fraction Unobserved is the fraction of nucleotides in the molecule which are not observed in the structure, compared to the longest experimental sequence in the EC.]

Popover shows structure quality details



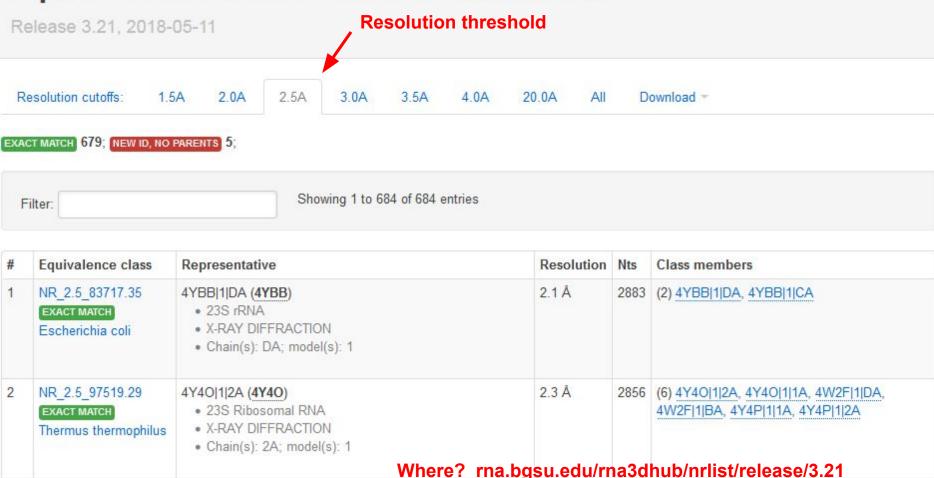
Where? rna.bgsu.edu/rna3dhub/nrlist/release/3.21

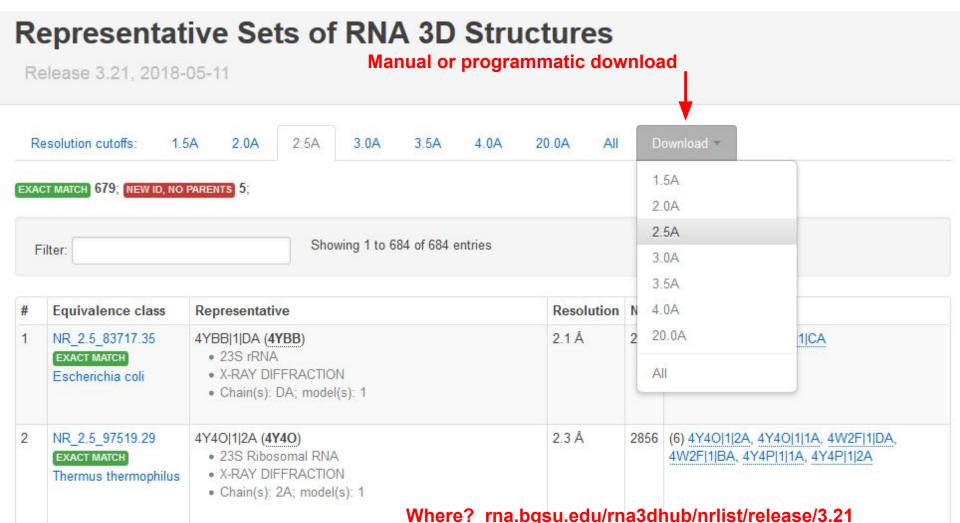
Representative set - take one IFE from each class

#	Equivalence class	Representative	Resolution	Nts	Class members
10	NR_4.0_75645.1 EXACT MATCH Toxoplasma gondii	5XXB 1 1+5XXB 1 4 (5XXB) 25S RNA 5 8S RNA ELECTRON MICROSCOPY Chain(s): 1, 4; model(s): 1	3.2 Å	3077	(1) 5XXB 1 1+5XXB 1 4
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Where? rna.bgsu.edu/rna3dhub/nrlist/release/3.21

Representative Sets of RNA 3D Structures





Annotations of all 3D structures

RNA Structure Atlas

- provides annotations of base-pairing, -stacking, and -backbone interactions computed by FR3D
- extracts internal, hairpin and 3-way junction loops and annotates them with motifs from the RNA 3D Motif
- organizes redundant structures into representative sets

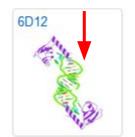
Choose one of 9862 RNA-containing 3D structures ▼

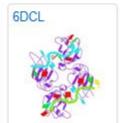
Recent Structures









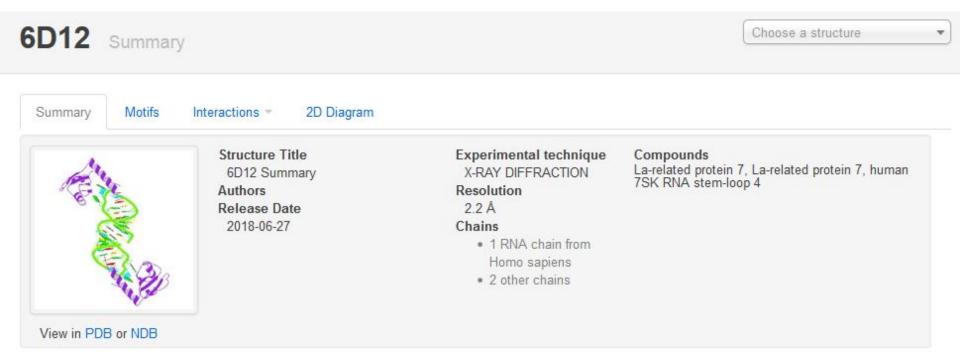






Where? rna.bgsu.edu/rna3dhub/pdb

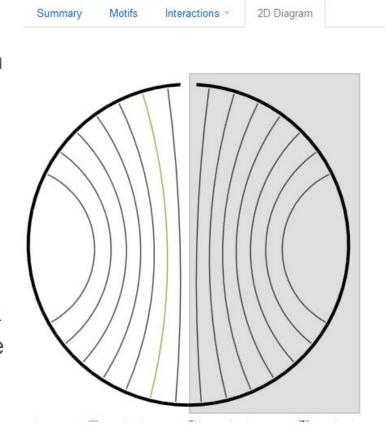
New structure 6D12

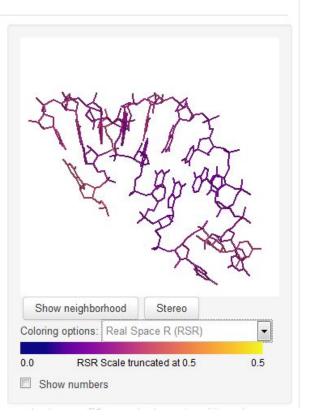


New structure 6D12, circular basepair diagram, RSR

Double helix with one non-WC basepair (green)

Selected
nucleotides
shown in 3D
window, colored
by Real Space R
(RSR) to indicate
structure quality





Where? rna.bgsu.edu/rna3dhub/pdb/6D12

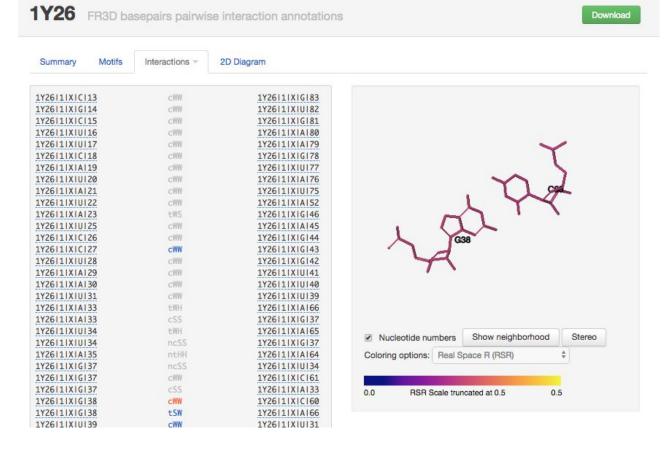
RSR/Z Viewer

We have added a new feature to the 3D Viewer in the BGSU RNA pages that allows users to color residues by RSR and RSRZ values.

The list shows all basepairs in structure 1Y26

Each basepair can be visualized, colored by RSR value.

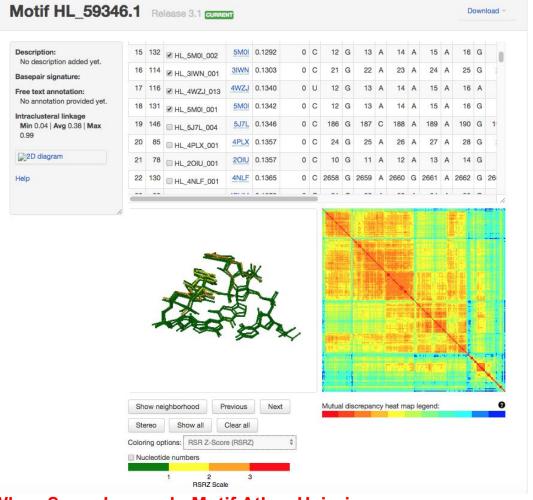
Basepair classification limits updated in 2017.



Where? rna.bgsu.edu/rna3dhub/pdb/1Y26

RNA 3D Motif Atlas and RSRZ coloring

Superposed instances of a hairpin loop motif group, colored by RSRZ, using the colorbar shown below the coordinates.



Where? rna.bgsu.edu Motif Atlas, Hairpin

Real Space R (RSR) & RSRZ Server

To support the visualizations, we have built a coordinate server, which returns 3D coordinates of specified residues in PDB format, and an RSR/Z server, which returns RSR and RSRZ values of specified residues.

RSR example, specifying two nucleotides by their Unit ID

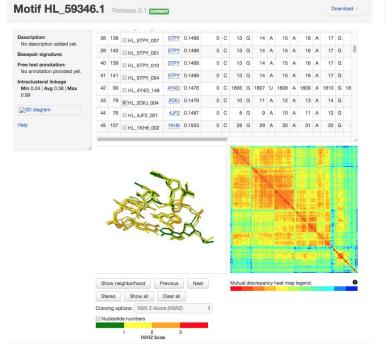
http://rna.bgsu.edu/rna3dhub/rest/getRSR?quality=1FJG|1|A|A|16,1FJG|1|A|C|18

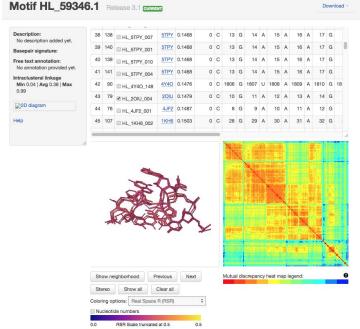
RSRZ example, specifying a hairpin loop by its Loop ID

http://rna.bgsu.edu/rna3dhub/rest/getRSRZ?quality=HL_1FJG_001

Applications of the RSR/Z Viewer

Coloring residues by RSR/RSRZ helps users identify interactions or parts of a motif that are not structurally 'reliable' or problematic. These are usually regions in a structure that are flexible or involved in local motion. This allows us to determine post-annotation steps that are required to ensure only interactions/motifs that have high structural confidence data are stored in our database.





Some instances in GNRA motif hiah have group RSR/Z average values. and might need to be filtered out in future releases of the Atlas Motif (HL 20IU 001, HL 20IU 004, HL 5F9F 003)

Representative Sets Generation

We present weekly releases (starting from 2011) of RNA structures grouped into sequence/structure equivalence classes on the BGSU RNA Site.

http://rna.bgsu.edu/rna3dhub/nrlist/

Each class includes a representative structure, originally selected as the structure with the most annotated base pairs per nucleotide. Starting with Release 3.0 (structures released through 15 December 2017), all representatives for equivalence classes were selected using Composite Quality Scoring (CQS). We are steadily building the missing releases at the rate of 2-3 per week; the current release is 3.21, structures released through 11 May 2018.