# Regulation of Gene Expression via Unproductive Splicing 

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# Integrative transcriptomic analysis suggests new autoregulatory splicing events coupled with nonsense-mediated mRNA decay 

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## ABSTRACT

Nonsense-mediated decay (NMD) is a eukaryotic mRNA surveillance system that selectively
is maintained by a large number of protein factors and cis-regulatory elements, which control the balance between mRNA production and degradation ( 1,2 ). Nonsense mutations and frame-shifting splicing errors induce premature

## Nonsense-mediated mRNA decay ${ }^{1}$

mRNA
 arana
 $\swarrow$ Splicing


Normal transcript


Protein production

${ }^{1}$ Kurosaki T, Maquat LE., Nonsense-mediated mRNA decay in humans at a glance., J Cell Sci. 2016 129(3):461-7

## AS-NMD Events Associated With Ultraconserved DNA Elements²



- AS-NMD is in every member of the human SR family
- Poison exons have evolved independently in most SR genes

[^0]
## Autoregulation of RBP by Nonsense-Mediated Decay (NMD)

- Poison exons cause NMD when included
- Essential exons cause NMD when skipped
- Exons 6 and 12 of RBM10 gene are essential ${ }^{3}$


[^1]
## Can we identify autoregulatory feedback loops?

- Inactivation of NMD $\rightarrow$ poison and essential exons ${ }^{1}$
- RBP perturbation followed by RNA-seq $\rightarrow$ regulated exons ${ }^{2}$
- CLIP $\rightarrow$ RBP binding to RNA ${ }^{2}$

[^2]
## Percent-Spliced-In (PSI, $\Psi$ )



PSI $=\Psi \simeq$ proportion of transcripts

$$
\psi=\frac{i n c}{i n c+e x c}
$$

$S J=i n c+e x c \simeq$ local expression level

## Statistical Significance of $\psi$



$$
\begin{aligned}
& \Delta \psi=\beta_{0}+\beta_{1} \log _{10}(S J)+e_{i} \rightarrow \text { residuals } \\
& z=\frac{\Delta \psi-\mu(\mathrm{SJ})}{\sigma(\mathrm{SJ})} \rightarrow \mathrm{p} \text {-value } \rightarrow \mathrm{q} \text {-value }
\end{aligned}
$$

## Splicing Factors Respond to UPF1/XRN1 Co-depletion

$$
\Delta \Psi=\Psi(K D)-\Psi(\text { Control })
$$



## Poison exons are more included upon NMD inactivation



## Essential exons are more skipped upon NMD inactivation



Essential

$\Delta \Psi<0$


## NMD inactivation, depletion of host gene, and eCLIP



## NMD inactivation, depletion of host gene, and eCLIP



## Serine And Arginine Rich Splicing Factor 7 (SRSF7)



- Splicing factor important for nuclear export and translation
- Overexpressed in colon and lung cancer tissues
- SRSF7 knockdown promotes apoptosis of colon and lung cancer cells
- SRSF7 regulates the splicing of the apoptosis regulator Fas
- SRSF7 maintains its homeostasis through the expression of Split-ORFs and nuclear body assembly (Königs et al, Nat Struct Mol Biol. 2020 Mar;27(3):260-273)
bioRxiv posts many COVID19-related papers. A reminder: they have not been formally peer-reviewed and should not guide health-related behavior or be reported in the press as conclusive.


## New Results

## Tissue-specific regulation of gene expression via unproductive splicing

Alexey Mironov, Maria Vlasenok, Sergei Margasyuk, Andrei A. Mironov, © Dmitri D. Pervouchine doi: https://doi.org/10.1101/2022.07.03.498634

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$\square$
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## Abstract

Eukaryotic gene expression is regulated post-transcriptionally by a universal mechanism called


GTEX

## Unproductive Splicing Events (USE)



## Validated Unproductive Splicing Events



## Validated Unproductive Splicing Events in SR proteins



Association btw. NMD isoform

## and gene expression:

not significant
significant

- NMD-promoting $\rightarrow$ NMD-inhibiting
mutual regulation: $\quad \square \quad \leftarrow-1$
— evidence of binding ..... no evidence


## Validated Unproductive Splicing Events in GTEx

Estimating changes in gene expression between the upper and the lower quartile of $\Psi$ distribution using Mann-Whitney U-test


## Negative association between $\Psi$ and host gene expression level


$\Psi_{H}=$ median of the upper quartile
$\Psi_{L}=$ median of the lower quartile
$\Delta e_{l}=$ gene expression change (local)
$\Delta e_{g}=$ gene expression change (global)
$z=z$-score of Mann-Whitney test for $e_{g}$

## Prediction of regulation by RBP

- Association of $\Psi$ and $e_{g}$ in GTEx: unproductive splicing
- Response of $\Psi$ to RBP perturbations: potential regulators
- Association of RBP expression and $e_{g}$ in GTEx: candidate regulators
- Additional evidence from CLIP, proteomics data etc

| Unproductive Splicing | Validated | Novel | Total |
| :--- | ---: | ---: | ---: |
| All | 48 | 2,831 | 2,879 |
| Significant | 11 | 568 | 579 |
| Tissue-specific | 5 | 86 | 91 |
| Regulated | 3 | 47 | 50 |
| CLIP in the gene | 3 | 31 | 34 |
| Local CLIP support | 3 | 14 | 17 |

## Brain-specific expression of GABBR1

GABBR1


$-2.5$

$\begin{array}{ccc}-2.5 & 0.0 & 2.5 \\ \text { PTBP1 }\end{array}$
NMD-prom.
trend test: NS


- 1. adipose - visceral
- 2. adrenal gland
- 3. artery - aorta
- 4. artery - tibial
- 5. bladder
- 6. brain
- 7. brain - amygdala
- 8. brain - anterior cingulate cortex

9. brain - caudate

- 10. brain - cerebellar hemisphere
- 20. colon

20. colon

- 11. brain - cerebellum
- 21. colon - sigmoid
- 30. lung
- 12. brain-cortex 22. colon-transverse
- 23. esophagus
- 31. ovary
- 13. brain - hippocampus
- 32. pituitary
- 14. brain - hypothalamus
- 24. esophagus - gastroesophageal

33. prostate

- 15. brain - nucleus accumbens - 25. esophagus - mucosa
$\begin{array}{ll}\text { - 16. brain - putamen } & \text { - 26. heart } \\ \text { - 17. brain - spinal cord } & \text { - 27. heart - atrial appendage }\end{array}$
$\begin{array}{ll}\text { - 16. brain - putamen } & \text { - 26. heart } \\ \text { - 17. brain - spinal cord } & \text { - 27. heart - atrial appendage }\end{array}$
- 28. heart - left ventricle
- 34. skin - sun exposed

17. brain - spinal cord

- 18. brain - substantia nigra
- 29. kidney
- 30. lung
- 35. small intestine - terminal
- 36. stomach
- 37. thyroid
- 38. uterus


## Predicted network of unproductive splicing events


cluster 1: increased $\Psi$ in the brain
cluster 2: increased $\Psi$ in the muscle
cluster 3: decreased $\Psi$ in the brain

## Brain-specific expression of DCLK4 is regulated by PTBP1 (unpublished)



## BRD2: long-range RNA structure around poison exon (unpublished)



BRD2 isoforms after ASO1 treatment (PCR)


Fold change of ratio of isoforms (RT-qPCR)


Credits to Marina Petrova and Dmitry Skvortsov

## Summary

- Auto- and cross-regulatory networks of unproductive splicing can be identified using large panels of transcriptomic data
- Integrative analysis of transcriptomic data brings novel insights into the structure of regulatory unproductive splicing networks, i.e., identification of novel targets and regulators
- RNA structure is involved in unproductive splicing regulation, possibly mediating the connection between protein binding and alternative splicing
- Positive feedback loops?
- Many other questions. . .


## Acknowledgments




[^0]:    ${ }^{2}$ Lareau et al, Unproductive splicing of SR genes associated with highly conserved and ultraconserved DNA elements. Nature, 446(7138), 926-9.

[^1]:    ${ }^{3}$ Yue Sun et al. NAR 45(14): 8524-8540, 2017

[^2]:    ${ }^{1}$ Lykke-Andersen et al. Human NMD initiates widely by endonucleolysis and targets snoRNA host genes. Genes Dev. 2014
    ${ }^{2}$ Van Nostrand et al. A large-scale binding and functional map of human RNA-binding proteins. Nature 2020)

