



BlockBuster2D: chimera reads clustering

Hua-Ting Yao

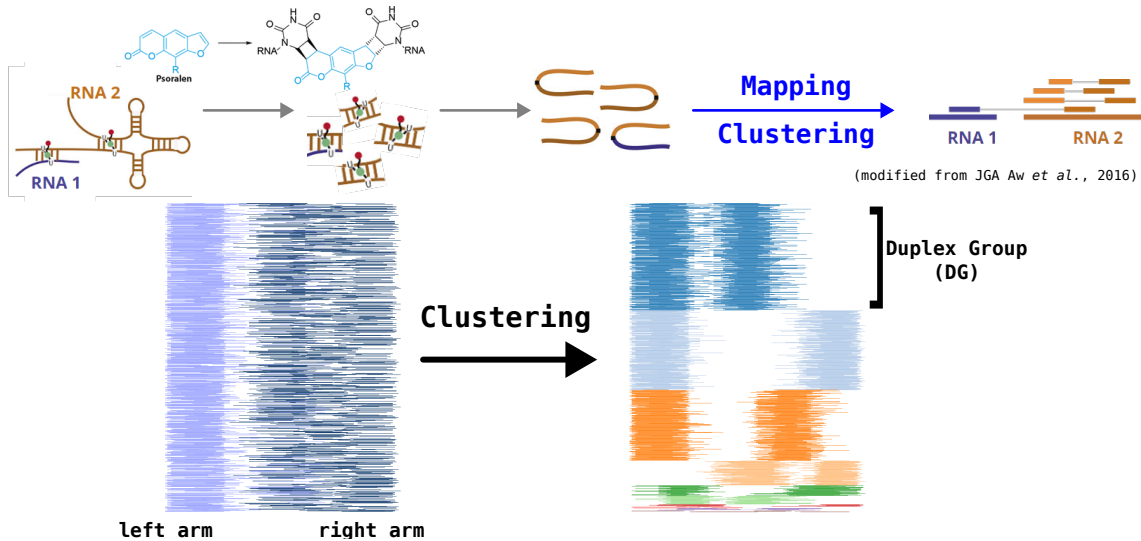
Theoretical Biochemistry Group (TBI), University of Vienna

Vienna, Austria

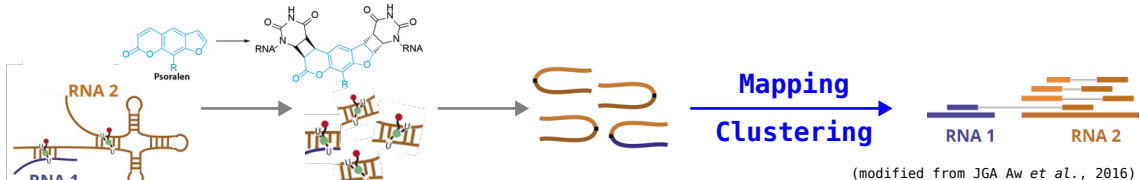


Katrin Gutenbrunner

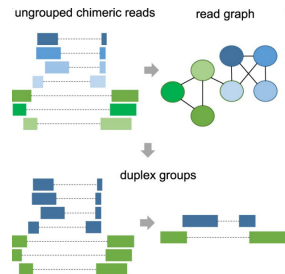
Cluster chimeric reads



Cluster chimeric reads

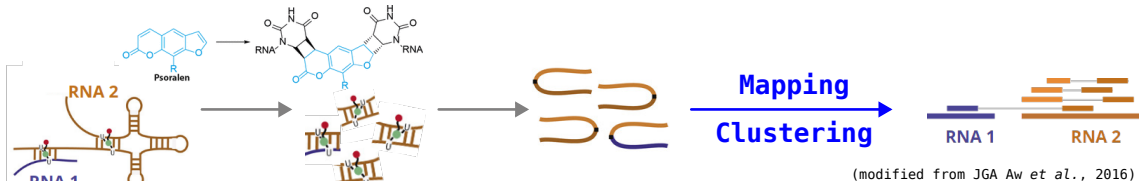


- Experimental protocols: [COMRADES](#), PARIS, SPLASH ...
- Clustering methods: CRSSANT, RANue, DuplexDiscoverer ...
- DG ranking: support read count, hybrid prediction ...

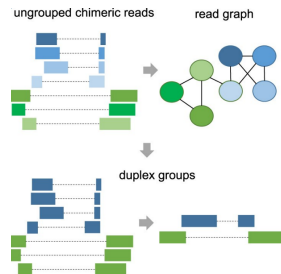


(E. Semchenko et al., 2025)

Cluster chimeric reads



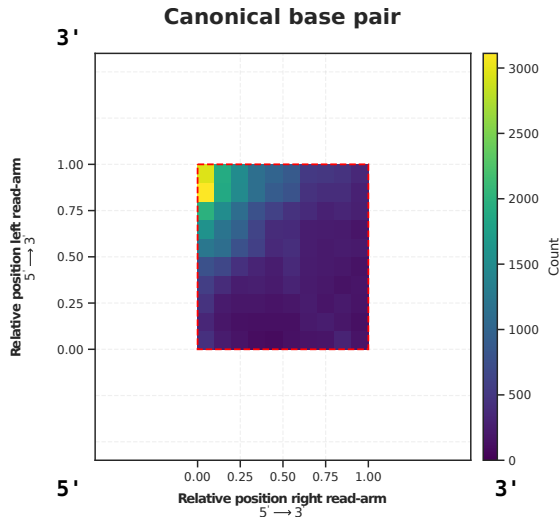
- Experimental protocols: [COMRADES](#), PARIS, SPLASH ...
- Clustering methods: CRSSANT, RANue, DuplexDiscoverer ...
- DG ranking: support read count, hybrid prediction ...



(E. Semchenko et al., 2025)

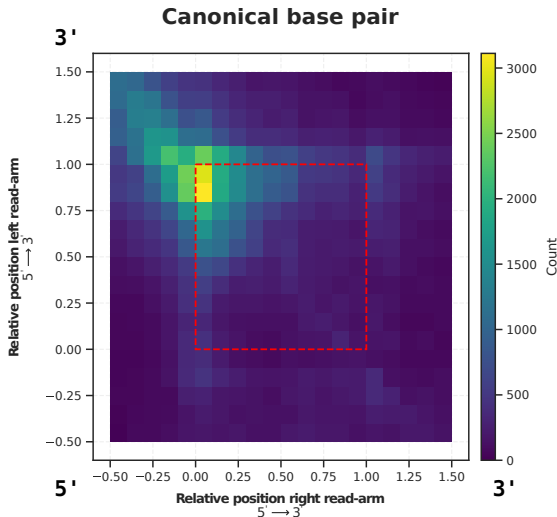
How to locate base pair in DG?

Base pair location in chimeric reads are strongly shifted (18s rRNA as example)



- 18s structure from 6YBS and 6YBW
- Found at least one canonical base pair in 24% chimeric reads

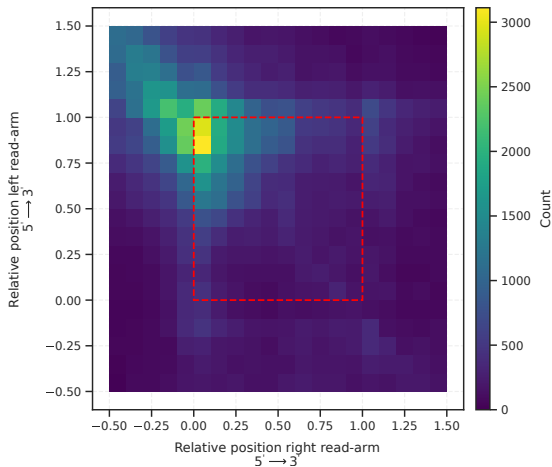
Base pair location in chimeric reads are strongly shifted (18s rRNA as example)



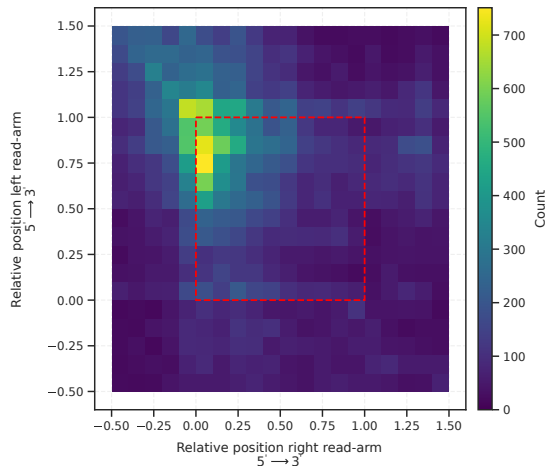
- 18s structure from 6YBS and 6YBW
- Found at least one canonical base pair in 24% chimeric reads
- Found at least one canonical base pair in 52% chimeric reads + extended regions

Base pair location in chimeric reads are strongly shifted (18s rRNA as example)

Canonical base pair

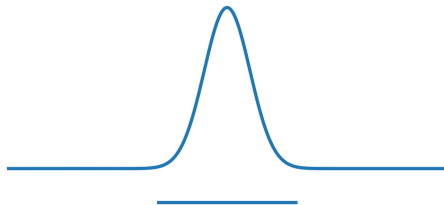


Non-canonical base pair



- 18s structure from 6YBS and 6YBW

- Identify miRNA (off)target blocks
- Each read is presented by a Gaussian distribution

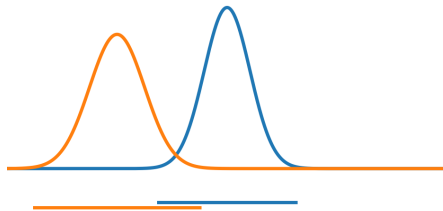


UGCCUAUUACCAGCGAAUGGACGUGUGGCGCACAGUUGAACGUAGACUAGAUAGUAUCAUAACGUAAACAGUAUGUACUAUGUAUCA

Reference

(D. Langenberger *et al.*, 2009)

- Identify miRNA (off)target blocks
- Each read is presented by a Gaussian distribution

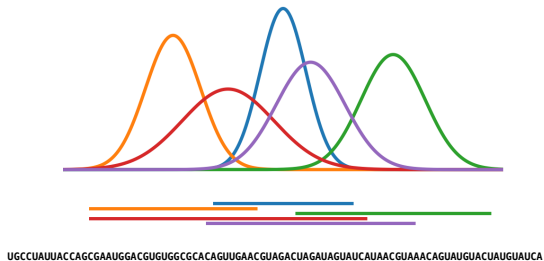


UGCCUAUUACCAGCGA AUGGACGUGUGGCGCACAGUUGAACGUAGACUAGAUAGUAUCAUAACGUAAACAGUAUGUACUAUGUAUCA

Reference

(D. Langenberger *et al.*, 2009)

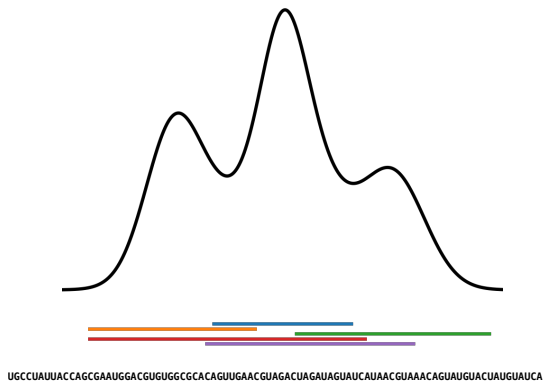
- Identify miRNA (off)target blocks
- Each read is presented by a Gaussian distribution



Reference

(D. Langenberger *et al.*, 2009)

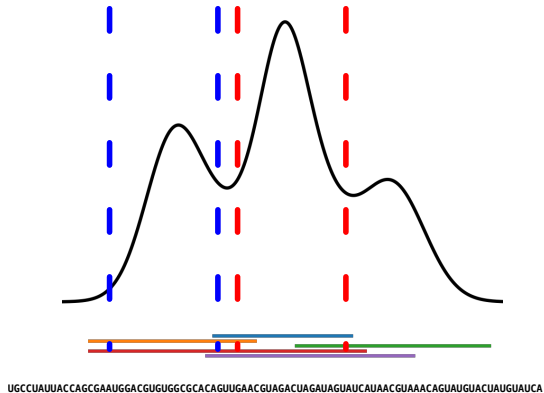
- Identify miRNA (off)target blocks
- Each read is presented by a Gaussian distribution



Reference

(D. Langenberger *et al.*, 2009)

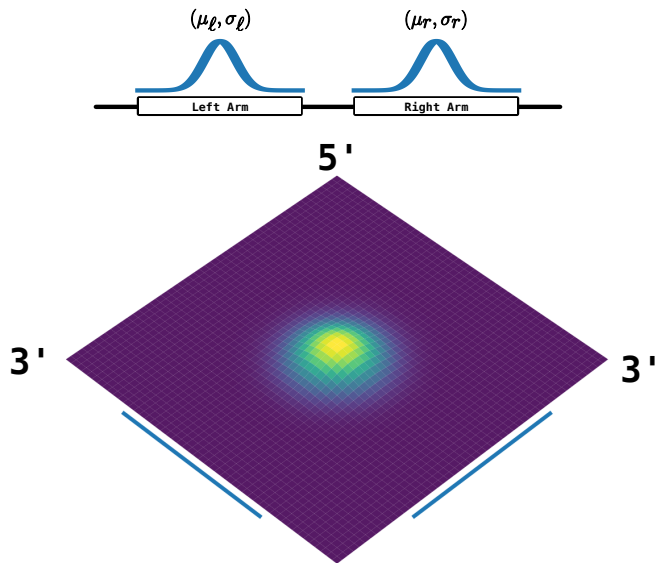
- Identify miRNA (off)target blocks
- Each read is presented by a Gaussian distribution



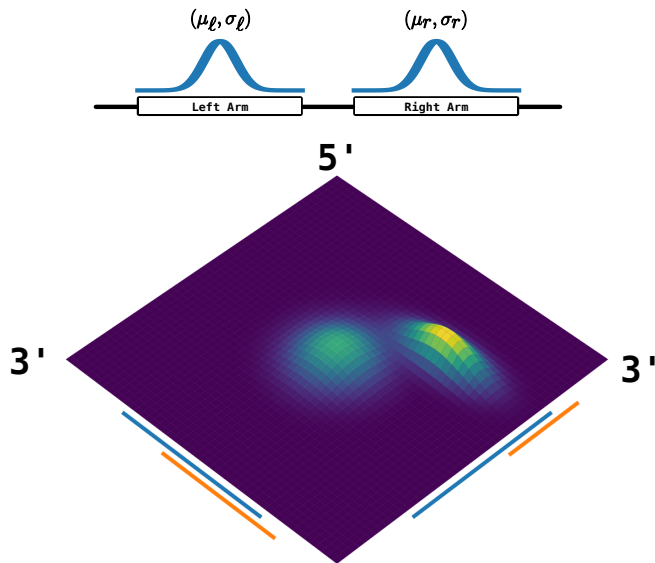
Reference

(D. Langenberger *et al.*, 2009)

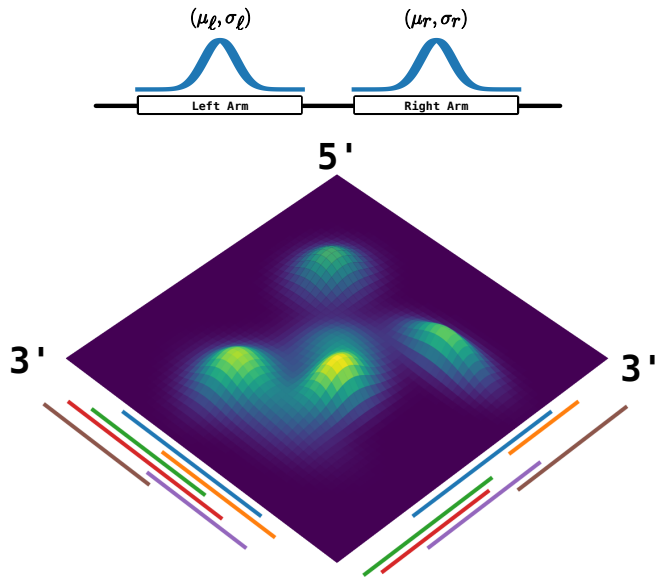
BlockBuster 2D: Let's add one dimension



BlockBuster 2D: Let's add one dimension



BlockBuster 2D: Let's add one dimension



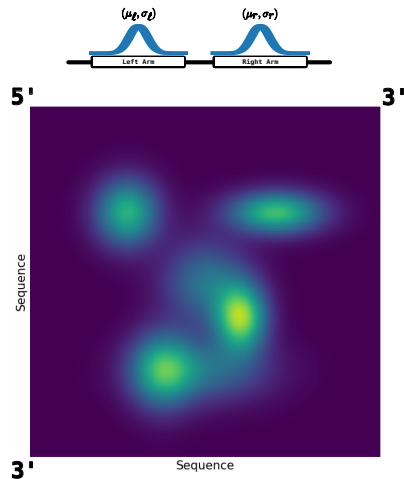
① Determine highest peak (p_x, p_y) from density function f

② Include in the block B , all chimeric reads (ℓ, r) s.t.

$$p_x \in [\mu_\ell - (\sigma_\ell + \delta_x), \mu_\ell + (\sigma_\ell + \delta_x)]$$

$$p_y \in [\mu_r - (\sigma_r + \delta_y), \mu_r + (\sigma_r + \delta_y)]$$

δ_x, δ_y : standard deviation of all μ_ℓ, μ_r added in B



① Determine highest peak (p_x, p_y) from density function f

② Include in the block B , all chimeric reads (ℓ, r) s.t.

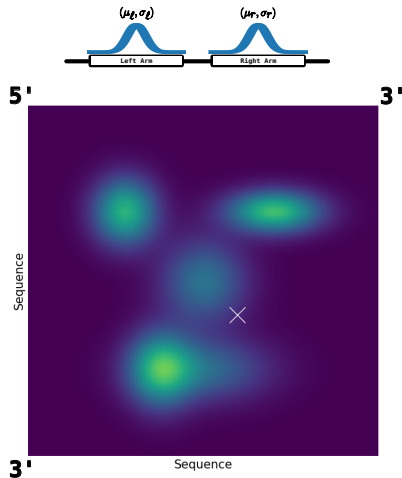
$$p_x \in [\mu_\ell - (\sigma_\ell + \delta_x), \mu_\ell + (\sigma_\ell + \delta_x)]$$

$$p_y \in [\mu_r - (\sigma_r + \delta_y), \mu_r + (\sigma_r + \delta_y)]$$

δ_x, δ_y : standard deviation of all μ_ℓ, μ_r added in B

③ Update density function $f \leftarrow f - f_B$

④ Repeat till $f = 0$



① Determine highest peak (p_x, p_y) from density function f

② Include in the block B , all chimeric reads (ℓ, r) s.t.

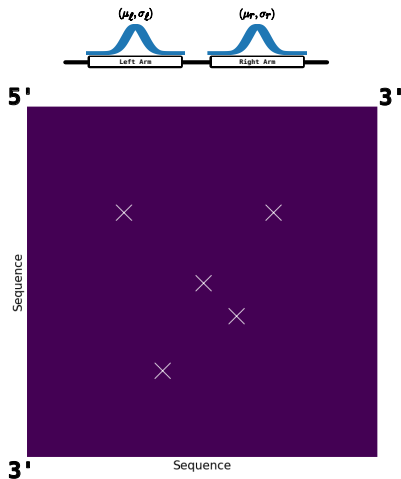
$$p_x \in [\mu_\ell - (\sigma_\ell + \delta_x), \mu_\ell + (\sigma_\ell + \delta_x)]$$

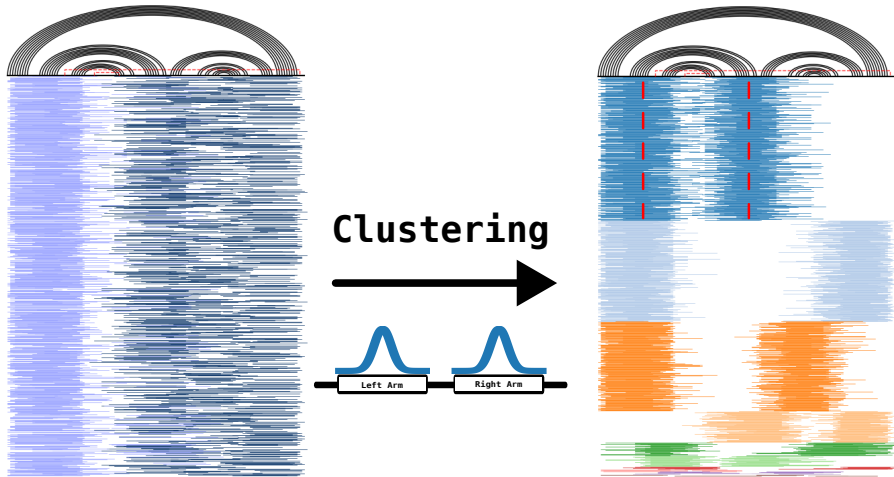
$$p_y \in [\mu_r - (\sigma_r + \delta_y), \mu_r + (\sigma_r + \delta_y)]$$

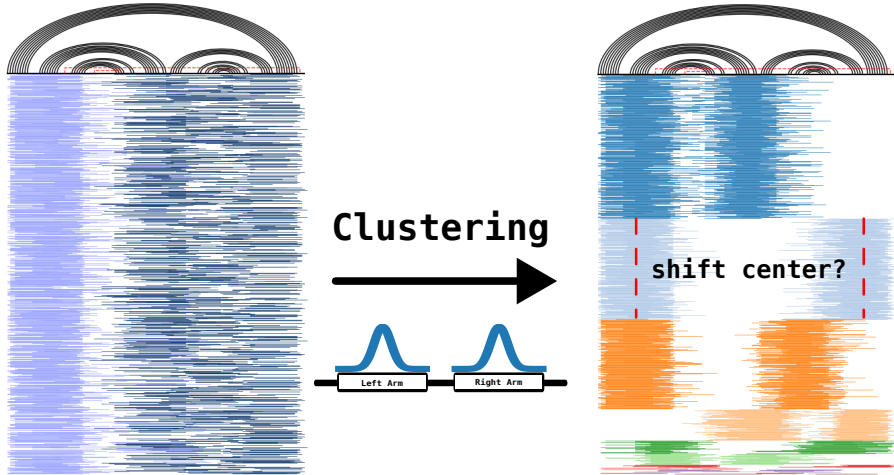
δ_x, δ_y : standard deviation of all μ_ℓ, μ_r added in B

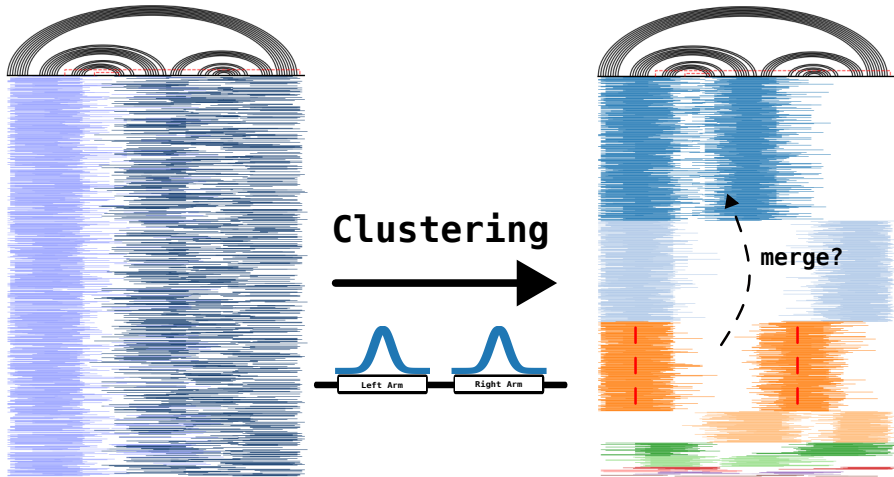
③ Update density function $f \leftarrow f - f_B$

④ Repeat till $f = 0$

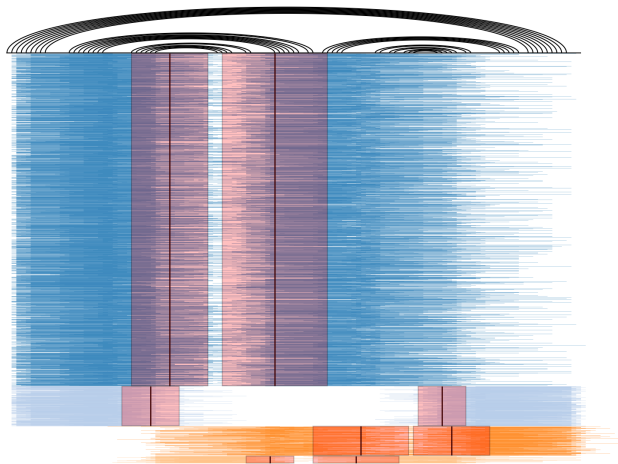
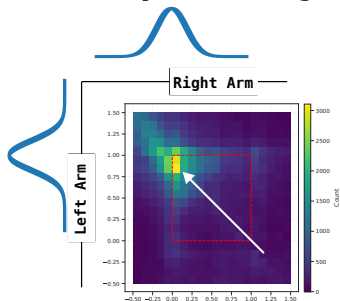






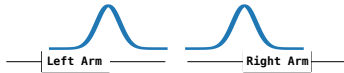


Density shifting

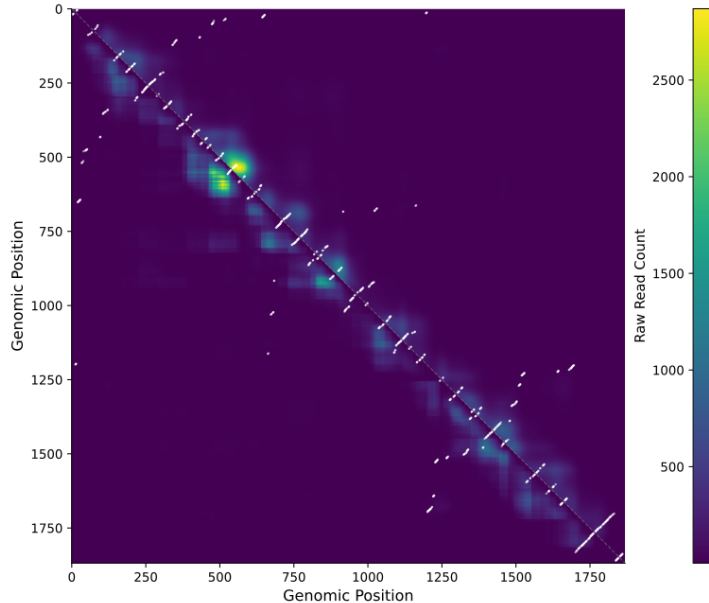


Density shifting solves some problems, but not all . . .

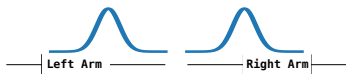
Case study: 18s RNA



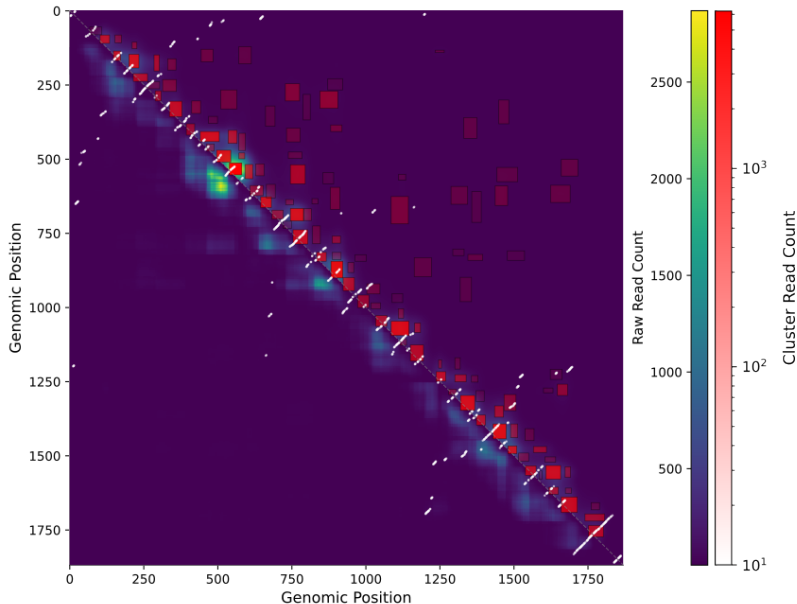
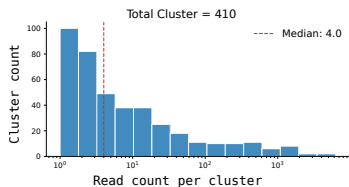
- reference (6YBS & 6YBW)
- white: canonical base pairs
- lower: Chimeric reads heatmap
- upper: Read density function



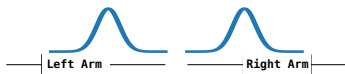
Case study: 18s RNA



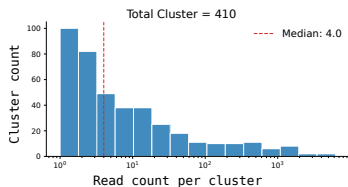
- reference (6YBS & 6YBW)
- white: canonical base pairs
- lower: Chimeric reads heatmap
- upper: **Clustered DGs**



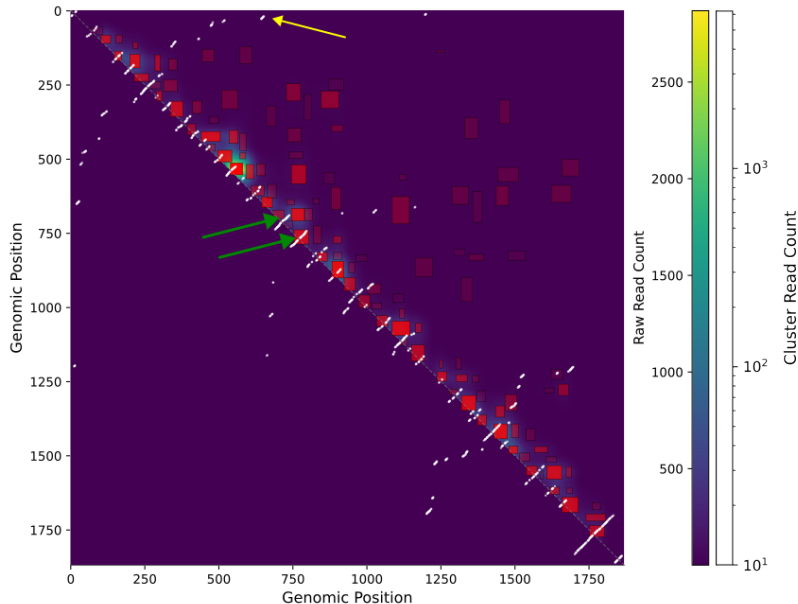
Case study: 18s RNA



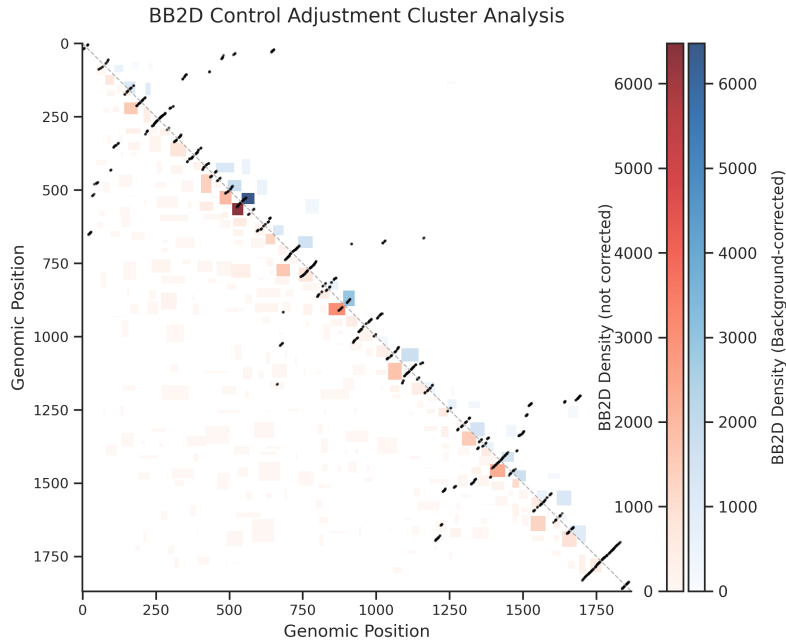
- reference (6YBS & 6YBW)
- white: canonical base pairs
- lower: Chimeric reads heatmap
- upper: **Clustered DGs**



Structure prediction is still
challenging



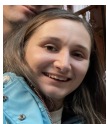
Case study: 18s RNA (Background noise correction)



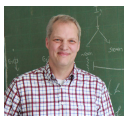
- Base pair location is bias
 - Protocol dependent?
 - How to locating base pair from crosslinking data
- Base pair is close to resulting duplex group
 - Can using empirical base pair distribution help?
 - Still need some effort for parameter calibration for fine tuned analysis
- Extension for PK identification and RNA-RNA interaction
 - combined with RNAnue or DuplexDiscoverer for fast screening



Katrin Gutenbrunner



Paulína Holotová



Ivo Hofacker

Denis Skibinski

Thomas Spicher

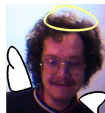
Leonhard Sidl

Yingjie Pan

Maximilian Faissner

Cristian A. Velandia-Huerto

⋮



Peter Stadler

Sarah von Löhneysen

Maria Waldl

Ronny Lorenz

⋮



Mimi & Maomao